Effects of Oral Administration of Red Palm Oil on Dichlorvos Induced Testicular Toxicity and Sperm Parameters in Albino Rats

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Authors’ contributions

This work was carried out in collaboration among all authors. Authors IKL, ADO and MTA were responsible for study designed and supervision of the research. Authors IKL and MTA performed the statistical analysis and wrote the protocol. Authors IKL and ADO wrote the first draft of the manuscript. Authors AAA and OAO managed the analyses of the study. Author IOA managed the literature searches. All authors read and approved the final manuscript.

ABSTRACT

Dichlorvos (2, 2-Dichlorovinyl Dimethyl Phosphate also known as DDVP) is an an organophosphate pesticide/ insecticide having oncogenic, genotoxic, neurotoxic and reproductive toxicity effects on the body. Elaeis guineensis (Palm oil) is an edible vegetable oil derived from the mesocarp ( reddish pulp) of fruit of the oil with antioxidant properties, beta-carotene content with highly saturated vegetable fats and is semi-solid at room temperature. This study was carried out to investigate the effects of oral administration of Red Palm Oil (RPO) on Dichlorvos (DDVP) induced testicular toxicity and sperm parameters in male albino rats. Concentration of 1ml of DDVP was diluted in 1000mls of distilled water and kept in a bottle for 3 days. 2 mls of red palm oil, 2 mls of

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INTRODUCTION

_Elaeis guineensis_ (palm oil) originated from West Africa and derived from mesocarp of the oil palm. It is widely used as cooking oil, basis of soap products (Unilever), American palm olive brand and industrial lubricant for machinery during British industrial revolution [1]. Palm oil is a perennial, evergreen tree adapted to cultivation in biodiversity rich equatorial land areas [2] and it is classified to be the most productive oil crop alone capable of fulfilling large growing world demand estimated to about 240 million tons by 2050 [3,4]. Palm oil is known to be the richest natural plant source of carotenoids in term of pro-vitamin A (retinol) equivalents such as a-carotene and b-carotene [5]. Red palm oil (RPO) exerts effects on reproductive capacity by improving the efficiency of protein utilization or biosynthesis in order to boost and improve sex hormone function. Red palm oil provides vitamin A which is a major factor in reproduction through the synthesis of sexual steroids, embryogenesis and spermatogenesis ([6], Chandrasekharan et al., 2004). _Elaeis guineensis_ has been reported to have protective antioxidant properties and carotenoids present in it have protective effects in cellular ageing, atherosclerosis, cancer, arthritis and Alzheimer’s diseases (Nagendran et al., 2005). Presently, according to Who Health Organization (WHO) approximately 80% of the World’s population depends on indigenous or traditional medicine for their primary health needs (Panti et al., 2011)[7]. Red palm oil is distinctive from other plant and animal oil because it contain 50% saturated fatty acids, 40% unsaturated fatty acids and 10% polyunsaturated fatty acids [8].

_Dichlorvos_ (2, 2-dichlorovinyl dimethyl phosphate) is an organophosphate insecticides cum pesticides widely use to control household pests in developing countries [9]. It is traded under the names such as Dedeva, DDVP, Nogos, Nuvan, Sniper and Dasksh [10]. It was classified as 1B highly hazardous chemical [11]. The compound has been under controversy because of its prevalence in urban water ways and toxicity extends beyond insects [12,13]. DDVP is effective against amphids, mites, caterpillars, white flies, thrips and also for the treatment of parasitic worms infection in humans and animals. The major use of dichlorovos is for space treatment of food poisoning, handling and storage of plant, corral, holden pens, poultry houses, feed lots, commercial and institutional building [14]. The exposure of Dichlorvos has an effect on human health such as genotoxic, oncogenic, neurotoxic and reproductive effects [15]. However, the study on effects of DDVP on fertility of male rat via intra-peritoneal injection has significant decrease in sperm numbers and increase in sperm abnormalities [16]. Further study reported significant reduction in testosterone levels, distortion in the cells of seminiferous levels and hypertrophy of spermatogenial cell levels of adult rats fed with water contaminated with dichlorvos [17]. This study was designed specifically to attenuate/reduce/minimize the effects of Dichlorvos (DDVP) induced testicular toxicity and sperm parameters using red palm oil.

MATERIALS AND METHODS

2.1 Materials

Red palm oil, DDVP, Formal saline, Distilled water, Oral cannula, Needle and Syringe, Manual weighing machine, Sample bottles, Dissecting set, Gloves, Microscope, Feeding and drinking
trough, Glass slide, Normal saline, Bouin’s fluid, Alcohol, Hematoxylin, Chloroform, and Slab.

2.2 Animals and Materials

Eighteen (18) adult albino rats of average body weight between 125g and 250g were bought from Animal house section of Physiology Department Ladoke Akintola University of Technology, Ogbomoso, Oyo State, Nigeria. The animals were housed in clean polypropylene cages and maintained in animal house at constant 12h/12h dark and light cycle with strict adherence to the ethical procedure. The animals were acclimatized for 2 weeks (14 days). Red palm oil (RPO) with viscosity that can pass through oral cannula purchased from Oja Oosa Market Apomu, Osun State, Nigeria. Analytical grade Dichlorvos (DDVP) chemical was purchased from Agro ally store Ogbomoso, Oyo State Nigeria. Concentration of 1ml of DDVP was diluted in 1000mls of distilled water and kept in a bottle for 3 days.

2.3 Experimental Protocol

The rats were divided into three groups (n=6), labeled as group A, B, C and D. Group A represent Control and were given water only, Group B were orally administered with 2 ml of red palm oil, Group C orally received 1 ml/100 g of DDVP and Group D orally received 2 ml of red palm oil plus 1 ml/100 g of DDVP as shown in Table 1. Feed and water were made available. The oral administration lasted for 2 weeks (14days) with oral cannula. Twenty four hours (24hrs) after the last treatment, the animals were sacrificed by cervical dislocation after which the testis were remove into formal saline filled sample bottles and the epididymis were mince in prewarmed normal saline (37°C) for sperm assessment. The sperm counts were analyzed using microscope.

2.4 Semen Analysis

The sperm count was evaluated/examined using new improved Neubauer’s hemocytometer (Deep 1/10 mm, LABART, Germany). The procedure reported by Jegede et al. [18] was adopted for the evaluation of sperm motility and sperm morphology. The sperm morphology grading is done using 2 drops of Wall and Ellas stain and air dried with the aid of microscope to view the shape and structure of spermatozoa in order to distinguish if there are any defects. Four grading were examined, which are morphology grading, head defect, middle- piece defect and tail defect.

2.5 Testicular Analysis

The method describe by Akpantal, (2000) was adopted in which the organ were cut in a slab of about 0.5cm thick transversely and fixed in Bouin’s fluid for a day after which it was transferred to 70% alcohol for dehydration. The tissues were passed through 90% alcohol and chloroform for different durations before been transferred into moisten paraffin wax for 20minutes each in an oven at 57°C. Serial sections were cut at 5microns and the slides were prepared from the tissues. The slides were dewaxed and passed absolute alcohol (2 changes), 70% alcohol and then to water for 5minutes and the slides were then stained with hematoxylin.

2.6 Data Analysis

Data obtained were analyzed using Microsoft Excel and results were expressed as mean +- standard deviation.

3. RESULTS AND DISCUSSION

3.1 Results

3.1.1 Physical appearance

No clinical signs indicative of systemic toxicity were observed in any animal during the study. The untreated rats appeared to be stronger, agile, well-fed and much better than the treated group which appeared slightly skinny, haggard, and aggressive. These changes are as a result of RPO and DDVP administered into the body of the rats. The results recorded for sperm count, sperm motility, morphology and testis histological analysis were presented below using each respective figure.

3.2 Discussion

3.2.1 Evaluation of sperm count

Sperm count analysis of the rats from all the groups were summarized in Fig. 1. The results obtained for sperm count showed that DDVP administration significantly reduced sperm count (p<0.05). There was no significant difference in the value obtained for RPO (Group B) induced rats in sperm count when compared with the
value of the control (Group A). However, the group administered with RPO + DDVP (Group C) indicates significant increase in sperm counts.

3.2.2 Evaluation of sperm motility and morphology

The sperm motility and morphological analyses showed significant changes in all motion parameters of rats as shown in Fig. 1. The study showed that DDVP administration significantly reduced both sperm motility and sperm morphology (p<0.05). There is significant decrease in the values recorded for RPO (Group B) as well as RPO + DDVP (Group C) administration when compared with the control (Group A). The seminal fluid assessments (sperm motility and sperm morphology) were significantly decreased in the treated groups (B, C, and D) compared with Group A. Similar observations were recorded in the work of Jegede et al. [19]. This study shows that the use of DDVP toxicity may cause oligospermia if administered in large volume for a long period of time. However, the motility and morphological values recorded for RPO administration is significantly higher compared to the values recorded for DDVP only and co-administered with RPO + DDVP. This agrees with previous report that palm oil is a potential supplement to increase fertility [20].

3.2.3 Histological analysis of the testis

In the present study, results indicates significant decline in sperm count, sperm motility and sperm morphology of the rats exposed to DDVP. Slightly increase in sperm count of the rat exposed to co-administered with RPO + DDVP but there is no significant difference in sperm count of the rats exposed to RPO. Cross section of histological analysis of the testis were presented in Fig. 2 for Group A, Fig. 3 for Group B, Fig. 4 for Group C and Fig. 5 for Group D respectively.

Group A: shows normal seminiferous tubules, spermatogenic cells and interstitial cells as shown in Fig. 2. According to the result of the analysis normal animal possess normal seminiferous tubules, spermatogenic cells and interstitial cells. It is evident that Group A was not administered with any of the substances and this serves as the control.

Group B: shows increase and well-formed seminiferous tubules with increase spermatogenic cells, Fig. 3. RPO (Elaeis guinensis) is known to be the richest natural plant source of carotenoids in terms of pro-vitamin A equivalent [21]. Vitamin A is known to play a part in reproduction through synthesis of sexual steroids. Aboua et al. [22] reported that the intake of palm oil promote the male sperm functions thus promoting spermatogenesis and embryogenesis.

Group C shows an improving spermatogenic cells and normal seminiferous tubules, Fig. 4. From the analysis disruption of seminiferous tubules in testis and reduction in sperm were able to come back to normal as a result of the presence of RPO induced. This is evident that palm oil contains antioxidants that act against free radicals caused by pollutant and pesticide reported by Nagendran et al. [23].

Group D shows laminar vacuolation, cellular degeneration and disruptions of seminiferous tubules, Fig. 5. The disruptions are due to the effects of 2, 2-dichlorovinyl dimethyl phosphate (DDVP) as one of the toxic substances causing damage on testis during spermatogenesis and resulted to testicular damage. Dichlorvos has been described by various studies as one of the toxic substances that cause damages on testis during spermatogenesis. According to Amara et al. [24] Dichlorvos toxicity on testis causes circulatory failure due to vascular damage and decreased utilization of electrolyte by spermatogenic cells due to its competitive action on the testis. This may further weaken fertility, suppress libido and deteriorate semen quality and testicular degeneration in animals.

<table>
<thead>
<tr>
<th>Grouping</th>
<th>Number of Rats</th>
<th>RPO</th>
<th>diluted DDVP</th>
<th>Ordinary Water</th>
</tr>
</thead>
<tbody>
<tr>
<td>Group A</td>
<td>6</td>
<td>-</td>
<td>-</td>
<td>+</td>
</tr>
<tr>
<td>Group B</td>
<td>6</td>
<td>+</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Group C</td>
<td>6</td>
<td>+</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>Group D</td>
<td>6</td>
<td>-</td>
<td>+</td>
<td>-</td>
</tr>
</tbody>
</table>
Fig. 1. Graph of mean values for sperm parameters

Fig. 2. Cross section of testicular histology analysis of Group A rat

Fig. 3. Cross section of testicular histology analysis of Group B rat

Fig. 4. Cross section of testicular histology analysis of Group C rat
Fig. 5. Cross section of testicular histology analysis of Group D rat

4. CONCLUSION

In conclusion, the results showed that RPO recorded better results over DDVP. There is no significant difference in the value obtained for RPO induced rats in sperm count when compared with the value of the control. RPO histological analysis shows increase and well-formed seminiferous tubules with increase spermatogenic cells. However, the motility and morphological values recorded for RPO administration is significantly higher compared to the values recorded for DDVP. Therefore, RPO offers positive protection against DDVP induced testicular damages. This study establishes that RPO exhibit no adverse effect on sperm parameters and could afford a protection against DDVP induced testicular toxicity.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

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Peer-review history:
The peer review history for this paper can be accessed here:
http://www.sdiarticle4.com/review-history/64667